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## Review Article

# The control and measurement of 'CO<sub>2</sub>' during fermentations

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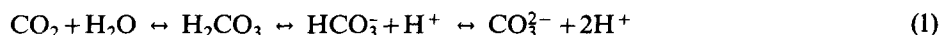
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## 1. Introduction

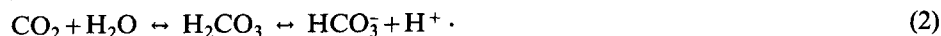
It has been known for many years that the growth and metabolism of microorganisms is accompanied by the uptake and/or evolution of CO<sub>2</sub>. To study and indeed to exploit the effects of CO<sub>2</sub> on microbial metabolism, it is necessary to control the level of (dissolved) CO<sub>2</sub> within the culture medium. Whilst several articles cover one or two methods by which *p*CO<sub>2</sub> may be measured and, hence, controlled, we know of no review of these. Thus, the purpose of the present article is to provide an overview of the role, measurement and control of the magnitude of *p*CO<sub>2</sub> during laboratory and industrial fermentations. We begin by describing the various 'CO<sub>2</sub>' equilibria and the question of CO<sub>2</sub>-absorption rates.

## 2. 'CO<sub>2</sub>' Concentrations

Although the partial pressure of CO<sub>2</sub> in the gas phase may be held constant, the ratios of the different possible species of 'CO<sub>2</sub>' in the aqueous phase will vary as a function of the pH and other factors. Since CO<sub>2</sub> can hydrate and dissociate in water, the reaction scheme may be written [1] as:



In addition, it has recently been proposed that small concentrations of dimeric hydrogen carbonate ions (H<sub>3</sub>C<sub>2</sub>O<sub>6</sub><sup>−</sup>) exist near neutral pH [2]. Since the concentration of this species is negligible, however, such ions will not be considered in the following. At pH values of < 8, the concentration of carbonate ions may be neglected [3] and only the following hydration reactions need to be considered:



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### 2.1. Dissolved CO<sub>2</sub> concentration

The concentration of CO<sub>2</sub> in solution ([CO<sub>2</sub>]<sub>aq</sub>) is normally expressed by Henry's law [4]:

$$[\text{CO}_2]_{\text{aq}} = K_H p\text{CO}_2 \quad (3)$$

where  $K_H$  = Henry's law constant (mol·atm<sup>-1</sup>) and  $p\text{CO}_2$  = the partial pressure of CO<sub>2</sub> in the gas phase (atm). For cultures grown under atmospheric pressure, the proportionality of solubility and partial pressures (Henry's law) may be assumed without introducing appreciable errors [5].

At a temperature of 37 °C,  $K_H = 10^{-1.61}$  [4] where the [CO<sub>2</sub>] is expressed in molar terms. Thus, to obtain [CO<sub>2</sub>] in millimolar terms,  $K_H = 10^{1.39}$ . Hence:

$$[\text{CO}_2]_{\text{aq}} = 10^{1.39} \times p\text{CO}_2 = 24.6 \times p\text{CO}_2.$$

In other words, when  $p\text{CO}_2 = 1$  atm, the concentration of dissolved CO<sub>2</sub> = 24.6 mmol·l<sup>-1</sup>.

### 2.2. Bicarbonate concentration in a pH-controlled culture with a constant temperature and gas phase

The equilibrium between CO<sub>2</sub> and HCO<sub>3</sub><sup>-</sup> is defined by a 'hybrid' equilibrium constant  $K_1'$  [4] where:

$$K_1' = \frac{10^{-\text{pH}} \cdot [\text{HCO}_3^-]}{[\text{CO}_2]} \quad (4)$$

From Eqn. 4, it follows that:

$$\log[\text{HCO}_3^-] = \text{pH} - \text{p}K_1' + \log[\text{CO}_2] \quad (5)$$

$\text{p}K_1'$  is related to the thermodynamic  $\text{p}K$  of the reaction  $\text{p}K_1^\circ$  and the ionic strength  $I$  by:

$$\text{p}K_1' = \text{p}K_1^\circ - 0.5f(I) - bI. \quad (6)$$

From Davies's Eqn. 4:

$$f(I) = [I^{1/2}/(1 + I^{1/2}) - 0.21][298/(T + 273)]^{2/3} \quad (7)$$

where  $T$  is the temperature in °C.  $I$  is the ionic strength of the medium and is given by [4, 6]:

$$I = \frac{1}{2} \sum c_i z_i^2 \quad (8)$$

where  $c_i$  = the concentration of ion  $i$  and  $z_i$  = the charge on ion  $i$ .

To obtain the apparent  $\text{p}K_{a,1}$  for the CO<sub>2</sub>/HCO<sub>3</sub><sup>-</sup> equilibrium, we use [4]:

$$pK_{a,1} = pK_{a,1}^{\circ} - f(I) - bI \quad (9)$$

Therefore, from Eqn. 9, we obtain  $pK_{a,1}$ , and so from Eqn. 5:

$$\log[\text{HCO}_3^-] = \text{pH} - pK_{a,1} + \log[\text{CO}_2].$$

It may be noted that these equations assume that there is an equilibrium between the  $p\text{CO}_2$  in the gas phase and that in solution. Clearly, this implies that the rate of exchange of  $\text{CO}_2$  between these two phases is rapid. It is, therefore, pertinent to determine the extent to which this is so.

### 3. Rates of Exchange of $\text{CO}_2$ between Fermentor Broths and Gas Phase

Following a step increase in the  $p\text{CO}_2$  in the gas phase, the rate of gas absorption ( $R_s$ )  $\cdot \text{vol}^{-1}$  of liquid is given [7] by:

$$R_s = \frac{dc}{dt} = \frac{k_L a}{h} (c_s - c) \quad (10)$$

where  $c_s$  = gas concentration at the interface,  $c$  = concentration of the gas in the bulk liquid,  $a$  = interfacial area,  $k_L$  = a constant dependent upon the diffusion coefficient of the gas and  $h$  = the thickness of the stationary film.

In general, because the value of  $h$  is not known, it is usual to combine it with  $k_L$  and write [6, 7]:

$$R_s = K_L a (c_s - c). \quad (11)$$

The absorption rate can be expressed in terms of partial pressures in gas and liquid ( $p_g$  and  $p_l$ , respectively) by making the substitutions  $c_s = H p_g$  and  $c = H p_l$  (where  $H$  = Henry's law constant) into Eqn. 11:

$$R_s = K_L a H (p_g - p_l). \quad (12)$$

The maximum absorption rate for a given partial pressure occurs when  $c=0$  and, hence,  $H p_l = 0$ . Then:

$$R_{s,\max} = K_L a c_s = K_L a H p_g. \quad (13)$$

The rate of absorption of  $\text{CO}_2$  is given by the slope of plots of ' $\text{CO}_2$ ' concentration vs. time. From Eqn. 13, it can be seen that:

$$K_L a = R_s / H p_g.$$

At  $37^\circ\text{C}$ ,  $H = 10^{-1.61}$  when ' $\text{CO}_2$ ' is expressed in M [4] and  $H = 10^{1.39}$  when ' $\text{CO}_2$ ' is expressed in mM. Hence:

$$K_L a = R_s / 10^{1.39} \cdot p_g.$$

For a detailed review of the diffusion, desorption and solubility of  $\text{CO}_2$ , in addition to some of the reactions involving  $\text{CO}_2$ , see also Ho et al. [6]. The rate of diffusion of  $\text{CO}_2$  through a silicone membrane may also be used to measure  $p\text{CO}_2$  [7a]. Having outlined  $\text{CO}_2$ -absorption rates and equilibria, we will now discuss various methods by which  $\text{CO}_2$  may be estimated (Table 1).

#### 4. Traditional Methods for Estimation of $\text{CO}_2$

The detection of  $\text{CO}_2$  in cultures includes both qualitative and quantitative methods. Production of  $\text{CO}_2$  from various carbohydrate substrata is a useful diagnostic tool in the identification of bacteria. Several methods for detecting  $\text{CO}_2$  production have been used, e.g., Durham tubes inserted into broth cultures [8], displacement of agar plugs [8, 9] and the appearance of cracks in agar medium [10]. The Eldredge tube [11], which is a device for trapping  $\text{CO}_2$  in barium hydroxide, can be used for both the qualitative (observation of a precipitate) and quantitative (recovery of the

TABLE 1

METHODS USED IN DETERMINATION OF  $\text{CO}_2$ . METHODS ARE LISTED IN THE ORDER THAT THEY APPEAR IN TEXT

Method	Principle	Selected references
Titrimetric	A standard alkaline solution is neutralised after absorbing $\text{CO}_2$	20
Colourimetric	Modification of colour of an indicator solution	21, 22
Gravimetric	Isolation and weighing of a compound of $\text{CO}_2$ (e.g., barium carbonate)	12, 20
Volumetric (Orsat)	Decrease in volume of a gas sample after chemical removal of $\text{CO}_2$	27
Volumetric (Warburg)	Increase headspace volume above an acidified sample	31
Manometric	Increase in pressure of headspace above an acidified sample	31
Katharometer	Differences in thermal conductivities of gases	27
Mass spectroscopy	Differences in mass to charge ratios of ions	35
IR	Absorption of IR radiation	27
Fibre optic probes	Changes in light transmittance of pH indicator	48
Chromatography	Differences in column retention times of gases	36
Potentiometric	Changes in pH induced by $\text{CO}_2$	50
Conductiometric	Changes in water conductivity caused by $\text{CO}_2$	70, 71
Amperometric	Flow of electric current due to reduction of $\text{CO}_2$	75
Piezoelectric	Change in resonant frequency due to adsorption of $\text{CO}_2$	103

precipitate) detection of  $\text{CO}_2$  [12]. However, these procedures depend upon the evolution of gaseous  $\text{CO}_2$  from the growth medium which does not always occur due to the high solubility of  $\text{CO}_2$  in water. The insertion of a red-hot loop into cultures resulted in a copious evolution of gas [13, 14] and this method of  $\text{CO}_2$  detection is known as the hot-loop method. Displacement of agar plugs is used in the hot-tube method which is more sensitive than the hot-loop method [15]. In the hot-tube method, an agar plug is displaced by the evolution of gaseous  $\text{CO}_2$  when the culture is heated to  $80^\circ\text{C}$ .

In the estimation of  $\text{CO}_2$  by the Conway microdiffusion method [16, 17],  $\text{CO}_2$  is liberated by acidifying the sample. There have been a number of designs of the apparatus for the microdiffusion method [16–18] but they are all based upon the standard cell illustrated in Fig.1.  $\text{CO}_2$ , liberated from the sample in the outer compartment, is allowed to diffuse to the central compartment where it is absorbed by a standard barium hydroxide solution [ $\text{KOH}$  or  $\text{NaOH}$  may be used instead of  $\text{Ba}(\text{OH})_2$ ] containing thymolphthalein indicator. The contents of the inner compartment are then titrated with a standard hydrochloric acid until the thymolphthalein indicator is just colourless ( $\text{pH} \approx 9.3$ ) at which point the excess of barium hydroxide is neutralised [16, 17].  $\text{CO}_2$  in a liquid sample may be estimated directly. Phenolphthalein is added to the sample which is then titrated with a standard sodium hydroxide solution. Free  $\text{CO}_2$  reacts with the sodium hydroxide to form sodium bicarbonate. Completion of the reaction is indicated by the development of the pink colour characteristic of phenolphthalein at the equivalence point of  $\text{pH} 8.3$  [19]. Titrimetric methods may also be used for the determination of  $\text{CO}_2$  in gases. The  $\text{CO}_2$  in the gas sample is absorbed into a standard barium hydroxide solution which, after the addition of phenolphthalein indicator, is titrated with standard hydrochloric acid [20].

Colourimetric methods, for the estimation of the  $\text{CO}_2$  concentration in a gas, do not require titration of the  $\text{CO}_2$ -absorbing solution. The colour intensity of a solution of phenolphthalein's sodium salt decreases as the concentration of  $\text{CO}_2$  increases [21, 22]. Similarly, the concentration of  $\text{CO}_2$  may be estimated by the modification of the colour of alizarin yellow R in the presence of  $\text{NaOH}$  [22].  $\text{CO}_2$ -detection tubes are also based on a colour change caused by a chemical reaction between the gas and tube

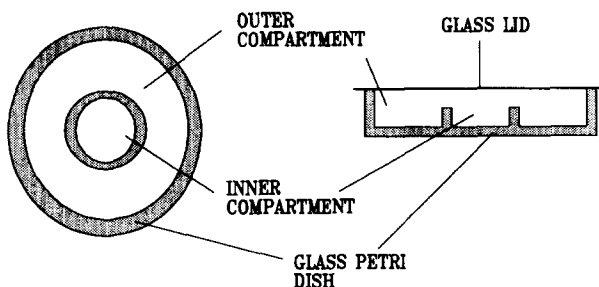


Fig. 1. Conway's standard microdiffusion cell. The cell consists of a Petri dish in the centre of which is fused a piece of glass tubing to form an inner cell or compartment. The edge of the outer dish is ground so that a lid, in the form of a square of flat ground glass, may enclose completely any gases within the dish. A mixture of vaseline and paraffin is smeared onto the lid to assist the seal.

contents [23]. When a gas containing  $\text{CO}_2$  is brought into equilibrium with sodium bicarbonate solution, the pH is a measure of the concentration of  $\text{CO}_2$  [24]. The pH may then be determined by the use of indicators [25] or with a pH electrode (see also Section 9.1.1.). When combined with a spectrophotometer the pH, and hence  $p\text{CO}_2$ , of a sodium bicarbonate solution, containing indicator and sparged with a  $\text{CO}_2$ -containing gas, may be estimated on a continuous basis [25].

Other traditional methods of measuring  $\text{CO}_2$  involve the passage of a measured stream of gas through absorbants followed by gravimetric [e.g., 12] or volumetric measurements [26, 27]. Such a method was employed in the Orsat apparatus in which the change in volume, at constant pressure, of a gaseous sample, following the removal of  $\text{CO}_2$  by its absorption into KOH, was measured and corresponded to the  $p\text{CO}_2$  [26, 27].

Total ' $\text{CO}_2$ ' may be measured by first saturating a sample with  $\text{CO}_2$ , then acidifying the sample and measuring either the change in the volume of the headspace at a constant pressure or the change in the pressure of the headspace at a constant volume [26, 28–32]. The latter method does not require gas meters for the accurate measurement of volume, and an example of an apparatus for such a method is shown in Fig. 2. Interference from volatile acids, which become gaseous in acidic conditions, is a possibility; however, interference due to organic or inorganic compounds commonly found in fermentation liquors are either negligible or easily eliminated [31]. Although such a method requires only very simple instrumentation and may easily be automated, it is not able to provide genuinely continuous monitoring of the total ' $\text{CO}_2$ '.

Enzymatic methods for the estimation of various compounds are specific for the compound(s) involved. One such method exists for the estimation of ' $\text{CO}_2$ ' and is commercially available (e.g., from the Sigma Chemical Company). The method is based upon the reaction catalysed by phosphoenolpyruvate carboxylase which produces oxaloacetate from phosphoenolpyruvate and ' $\text{CO}_2$ ' [33]. Malate dehydrogenase then

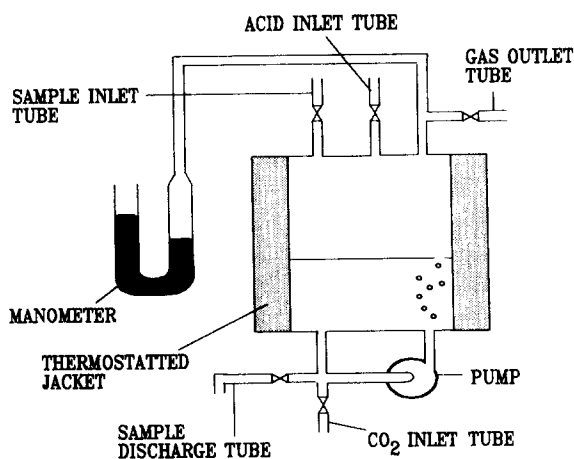


Fig. 2. Design of an apparatus for manometric determination of  $\text{CO}_2$ .  $\text{CO}_2$  is estimated by measuring the increase in pressure of the headspace following acidification of sample [31].

converts NADH and the so-formed oxaloacetate to malate and  $\text{NAD}^+$ . When the absorbance is read, at either 340 or 380 nm, the decrease in absorbance upon the addition of PEP is proportional to the original ' $\text{CO}_2$ ' content of the sample [33].

With the exception of the colourimetric method, the 'traditional' methods, discussed above, can only be used for spot checks and are therefore not suitable for continuous analysis. Methods employed in the gas analysis of clinical samples (e.g., blood) are also unsuitable for continuous use, since they are performed on samples taken from a patient [e.g., 32, 34].

## 5. $\text{CO}_2$ Analysis by Katharometer

The principle of  $\text{CO}_2$  analysis by katharometer [27] is as described below. The behaviour of the katharometer reflects the thermal conductivity of the gas (mixture) which is passing through it. Within the katharometer are four small cells, each containing a glass-coated Pt wire identical with the others. These four wires form the arms of a Wheatstone bridge (Fig. 3). Two of the cells (B, D) are exposed to a reference gas and the other two (A, C) are exposed to the sample gas. When the bridge current is constant and all four cells are exposed to the same gas, each wire will attain the same temperature and resistance. Under these conditions, the bridge is balanced and no current flows through the galvanometer.

If two gases of different thermal conductivities, such as air and an air/ $\text{CO}_2$  mixture, are introduced into the cells (air into the reference cells and the mixture into the sample cells), then Wires B and D will lose more heat than will Wires A and C. This is because the thermal conductivity of air is greater than that of  $\text{CO}_2$ . The consequent change in the conductivities of the wires will unbalance the bridge and cause a deflection of the galvanometer, with the size of the deflection being dependent upon the difference in the thermal conductivity, and in this instance  $p\text{CO}_2$ , of the two gases. However, the measurement of gases by katharometers is not specific, e.g., for  $\text{CO}_2$ . To measure the  $p\text{CO}_2$  of the effluent gas from a fermentor the constituent gases

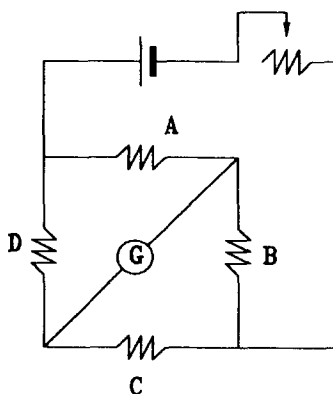


Fig. 3. A simplified representation of a katharometer. Sample and reference cells each contain a Pt wire (A–D), each of which is identical, forming the arms of a Wheatstone bridge. The principle of  $\text{CO}_2$  measurement using a katharometer is as described in text.

would first have to be separated, for example, by gas chromatography as described in Section 8.

## 6. Mass Spectrometry

The basic principle of mass spectrometry is the separation and registration of ionic masses [35]. The sample is ionised, e.g., by electron bombardment, and the ions, with a mass-to-charge ratio of  $m/z$ , are accelerated by an electric field through a series of slits to form an ion beam. This ion beam is then deflected by a magnetic field,  $H$ , according to  $m/z = (H^2 r^2)/2V$  where  $H$  is the strength of the magnetic field,  $r$  the curvature of the pathway (instrumentally dictated) and  $V$  the voltage used to accelerate the ions [35]. By continuous scanning of either the magnetic field or the accelerating voltage, ions of different  $m/z$  values can be focussed on the detector and subsequently measured. In addition to the identification of compounds, the mass spectrometer can be adapted for selected ion monitoring to permit highly selective determinations of very high sensitivity [36]. However, mass spectrometry cannot always be used to examine a number of substances simultaneously without additional separation techniques such as gas chromatography (see Section 1.5). The reason for this is that mass spectra tend to be complicated and examination of several compounds simultaneously can lead to significant overlap, seriously compromising the quantitation of the individual species [37]. Despite this problem, gases produced by fermentation processes can, in fact, be estimated simultaneously by mass spectrometry [38–40], by exploiting the fact that different gases have different isotopic compositions.  $\text{CO}_2$  is measured at  $m/z = 44$ . The only possible interference is from  $\text{N}_2\text{O}$  (formed by many denitrifiers) and from volatile fatty acids (which may break down to  $\text{CO}_2$  within the mass spectrometer [39]).

The type of mass spectrometer described above is the magnetic mass spectrometer which is perhaps more often used for the analysis of fermentor exhaust gases whilst quadrupole mass spectrometers are used for the estimation of dissolved gases [39]. These nonmagnetic mass spectrometers employ as a mass filter four electrically conducting parallel rods precisely located in a ceramic holder. The opposite rods are connected electrically and a radio-frequency ( $RF$ ) voltage superimposed on a  $DC$  voltage is applied to the two pairs of rods,  $180^\circ$  out of phase with each other [35, 38–40]. Ions entering at one end of this array are constrained to make oscillations in the transverse electric field. At a given  $RF/DC$  voltage, only ions of a specific  $m/z$  value will avoid collision with one of the rods and will, thus, pass through the filter. Scanning the  $RF$  and  $DC$  voltages may be performed rapidly with the ions being transmitted sequentially in order of their  $m/z$  ratios with constant resolution [35, 38–40].

Sampling dissolved gases is achieved by a membrane inlet to the mass spectrometer [39–41]. A perforated capillary or sheet, or porous material, is covered with a plastic membrane and immersed into the fermentation medium. A vacuum is applied to the inside of the probe and the dissolved gases diffuse through the membrane. Since the diffusion coefficient is dependent upon both the gas and the membrane used, the membrane inlet is selective. Another means by which the dissolved gases in fermentation media could be sampled is by a carrier gas tubing method [3, 42]. This method involves passing a carrier gas through a tube, or across a membrane, made of a gas-

porous material, immersed in the fermentation medium. The exhaust gas from the tubing is then analysed.

Unfortunately, methods such as mass spectrometry and infrared analyses (see the following section) are normally prohibitively expensive for routine use on single fermentors at the laboratory scale [43].

## 7. Optical Methods of CO<sub>2</sub> Determination

### 7.1. IR absorbance

The absorbance of IR radiation by a gas (mixture) is proportional to the mass of absorbing molecules in the light path and is, therefore, proportional to the absolute pressure of the gas in an analysis cell [27]. The principle of IR spectroscopy for estimating the partial pressures of gases is as follows [27, 44]. When analysing a single component in a gas stream, a so-called 'nondispersive analyser' is used. This examines absorbance in the wavelength range of 3–15  $\mu\text{m}$ . The light beam, after passing through the absorbance cell through which the gas is flowing, passes to a detector that is selective for the wavelength at which the component being analysed absorbs. The intensity of the transmitted light is compared with the intensity of the incident light by using a 'blank' or reference cell filled with CO<sub>2</sub>-free gas. The difference between the two signals is a measure of the  $p\text{CO}_2$ . In a null-balance instrument, this difference is used to actuate a servo system. This adjusts a shutter situated in the light path of the reference cell until the sample and reference radiations are matched. The shutter position is a measure of the  $p\text{CO}_2$ .

Water absorbs strongly in the IR [45] and, hence, the presence of water vapour in the gas sample may interfere with the IR spectrophotometric measurement of CO<sub>2</sub>. Since this absorption is characteristic of –OH bonds, volatile alcohols may also interfere. Some other compounds from bacterial cultures, which absorb in the IR and those may, thus, interfere, are acetone, acetaldehyde and substances with –SH bonds [44]. IR gas analysis is, therefore, not selective for CO<sub>2</sub> unless the gas sample has been treated to remove interfering substances prior to the  $p\text{CO}_2$  measurement and, as stated above, this technique may be rather expensive.

### 7.2. Fibre optic measurements

Stimulated by recent developments in fibre optics technology, a number of new uses of fibre optics in sensing have been proposed. The development of fibre optic pH probes [46] has led to the production of fibre optic CO<sub>2</sub> probes [47, 48]. In such fibre optic CO<sub>2</sub> probes, ambient  $p\text{CO}_2$  controls the pH of a bicarbonate buffer solution, contained within a membrane, which then influences the optical transmittance of a colourimetric pH indicator. One fibre carries light to the buffer/indicator solution, and a second fibre carries the transmitted signal (Fig. 4) to a receiver which converts the signal to an electrical output. The optical approach avoids problems of making miniature glass or liquid membrane  $p\text{CO}_2$  electrodes [49] with adequate reference electrode stability. However, the changes in pH, that are measured with such a device, are not necessarily due solely to a change in  $p\text{CO}_{2,\text{aq}}$  since organic acids or bases that are able to penetrate the membrane may also cause such a change in pH [50, 51].

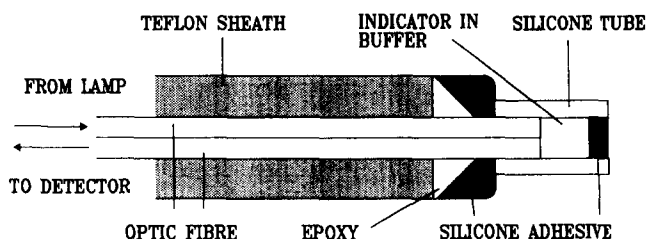


Fig. 4. Construction of sensor tip of a fibre optic probe. Two fibres extending from end of appropriately opaque and inert tubing are sealed into it with epoxy. Epoxy is also layered onto cut end of fibres to prevent binding of indicator dye. Clear silicone adhesive is coated over epoxy except at fibre ends. A piece of silicone tubing is then plugged at one end with white silicone adhesive and filled with a solution of phenol red,  $\text{KHCO}_3$  and  $\text{KCl}$  [48].

## 8. Chromatographic $\text{CO}_2$ Determination

In gas liquid chromatography, a sample is introduced into a stream of carrier gas (the mobile phase) and is swept through a chromatographic column which contains a stationary phase. After introduction onto the column, the sample components are allowed to distribute between the stationary and mobile phases according to their partition coefficients.  $K$ , the partition coefficient, is defined [36, 52] as:

$$K = C_s / C_m$$

where  $C_s$  is the concentration of solute in the stationary phase and  $C_m$  is the concentration of solute in the mobile phase. The lower the value of  $K$ , i.e., the lower the solubility of the solute in the stationary phase, the quicker it will pass through the column. Various components of a sample can be separated only if they have different  $K$  values. The time taken to pass through a column, the retention time, will be affected by the type of column, temperature and carrier-gas flow rate. Once separated, the components enter a detector (e.g., thermal conductivity detector, mass spectrometer) connected to a recorder or a printer/plotter and are consecutively registered as chromatographic peaks. Provided that the detector and separation procedure are appropriate, each peak will correspond to one of the components of the original sample. An internal standard may be added to the sample before analysis to compensate for possible variations in analytical conditions. The retention times of the components are then calculated relative to that of the internal standard. For a more detailed discussion of chromatographic separation theory, see, e.g., Larsson and Odham [36], Perry [52], Grob [53], Cramers and McNair [54] and Willett [55]. The area beneath each peak can then be correlated to the concentration of the corresponding component in the injected sample [36, 52, 54, 55]. However, since chromatographic techniques rely upon the spatial separation of the compounds that are being quantified, they are only of use on a noncontinuous basis.

## 9. CO<sub>2</sub> Electrodes

### 9.1. Potentiometric

Ion-selective electrodes may be utilised for the estimation of a wide range of compounds, including gases, e.g., ammonia, O<sub>2</sub> and CO<sub>2</sub> [56–58]. The determination of  $p\text{CO}_{2,\text{aq}}$  by ion-selective potentiometric electrodes, such as the Severinghaus-type (Fig. 5), involves the measurement of the pH of a thin layer of a buffer solution in equilibrium with ambient CO<sub>2</sub> [57, 59]. The fermentation medium and the buffer solution within the electrode are separated by a membrane which is 'freely' permeable to CO<sub>2</sub>. However, the use of the Severinghaus-type potentiometric electrode for measuring the levels of dissolved CO<sub>2</sub> is hampered by its slow response time; complex relationships among multiple chemical species during unsteady diffusion were shown to cause hysteresis and pH-dependent response rates [59]. Digital simulations may be used to predict the dynamic response of such electrodes [60] and electrode stability may be sacrificed to improve response time. As with the fibre optic probes mentioned above, the changes in pH that are measured with such a device are not necessarily due solely to a change in  $p\text{CO}_{2,\text{aq}}$  [50]; additionally, the electrodes are by no means cheap.

Another problem with devices, such as the Severinghaus-type electrode, is that they must be immersed in the fermentation broth and are, therefore, 'invasive'. The use of such invasive devices leads to a number of potential problems: an additional port on the fermentation vessel will be necessary leading to an increased risk of contamination: the device must be robust enough to withstand sterilisation: the device is required to be calibrated easily without having to be removed from the fermentation vessel. Such a steam-sterilizable Severinghaus-type  $p\text{CO}_2$  electrode that can be calibrated in situ, has, in fact, been described [61].

K<sub>2</sub>CO<sub>3</sub> may be used as a solid electrolyte for the potentiometric measurement of gaseous carbon oxides. It was shown that the triple contacts: K<sub>2</sub>CO<sub>3</sub>(s), CO<sub>2</sub>(g), Pt(s) and: K<sub>2</sub>CO<sub>3</sub>(s), CO<sub>2</sub>(g), ZrO<sub>2</sub>–CaO(s) can be used for CO<sub>2</sub> determination in air- or in O<sub>2</sub>-bearing gases at temperatures ranging from 450 to 750 °C [62].

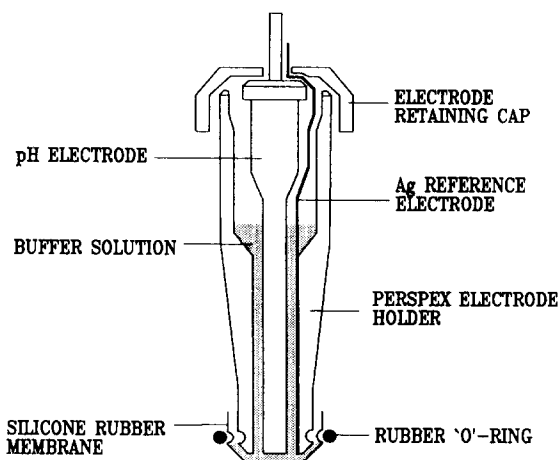


Fig. 5. Design of Severinghaus-type electrode. CO<sub>2</sub> is absorbed into or desorbs from buffer solution and corresponding change in pH is measured by pH electrode [50].

The fabrication and performance of a variety of ion-sensitive field-effect transistors (ISFET), which are in effect solid-state devices (no internal reference electrolyte), have been described [63–65]. The ISFET has attracted considerable interest because it is envisaged that a single miniaturised solid-state chip could contain multiple sensors and be used to sense several ions simultaneously. It has been shown that ISFETs based on ion-selective membranes were subject to positive interference by either  $\text{CO}_2$  or organic acids [51]. When field-effect transistors were coated with PVC or silicone rubber membranes, larger responses to  $\text{CO}_2$  than with ion-selective membranes were observed and the resulting ISFETs behaved very much like Severinghaus-type potentiometric  $\text{CO}_2$  sensors [51]. The response times, however, were much slower than with Severinghaus-type electrodes and the ISFETs still responded to organic acids. ISFETs, like Severinghaus-type electrodes, are invasive devices and, therefore, possess all of the problems associated with invasive devices listed above. However, for gas analysis, rather than the determination of the concentrations of particular ions in a solution, ISFETs could be used as noninvasive devices by simply placing them in the effluent gas flow.

*9.1.1. Invasive and noninvasive electrode methods for determination of  $\text{CO}_2$ .* The measurement of  $p\text{CO}_2$  in the effluent gas is said to give an excellent approximation of the  $p\text{CO}_{2,\text{aq}}$  [3, 66 but cf. 6], thus, eliminating the need for the more technically demanding measurement of  $p\text{CO}_{2,\text{aq}}$  and allowing the utilisation of noninvasive devices. It is possible, however, to use an invasive device in a noninvasive way. A potentiometric method was proposed whereby changes in pH, caused by the absorption of  $\text{CO}_2$  when the effluent gas was bubbled through a solution of sodium carbonate, were measured [23]. This method is still hampered by the problems associated with Severinghaus-type electrodes listed above. Noninvasive methods for determining  $p\text{CO}_2$  have received attention for clinical uses in the transcutaneous analysis of arterial  $p\text{CO}_2$  [e.g., 67–69]. The principle involved is once again the measurement of the pH change within a buffer solution and, despite the slow response time, such a technique is of use in control systems for maintaining a normal arterial  $p\text{CO}_2$  by artificial respiration since such sensors are capable of continuous measurement.

## 9.2. Conductimetric

Conductimetry has also been applied to the measurement of  $p\text{CO}_2$ . A continuous conductimetric sensor for  $\text{CO}_2$  (Fig. 6) was described in which  $\text{CO}_2$  diffused through a hydrophobic gas-porous membrane into a thin layer of pure water [70, 71]. The back wall of the thin water layer (see Fig. 6) was a porous screen separating the water from a mixed-bed ion-exchange column which continuously removed ionic species from the water layer. Electrodes positioned in the water layer continuously measured its conductance. The diffusion of  $\text{CO}_2$  into the water layer and the removal of ionic species by the mixed-bed ionic exchanger established a steady-state concentration gradient of  $\text{CO}_2$ . The gradient was proportional to  $p\text{CO}_2$  in the gas phase and the cell conductance was proportional to  $(p\text{CO}_2)^{1/2}$  [70, 71]. However, this device was evaluated for use in the determination of  $p\text{CO}_2$  only up to 0.01 atm.

## 9.3. Amperometric

In contrast to  $\text{CO}_2$ ,  $\text{O}_2$  is routinely and more-or-less reliably measured either

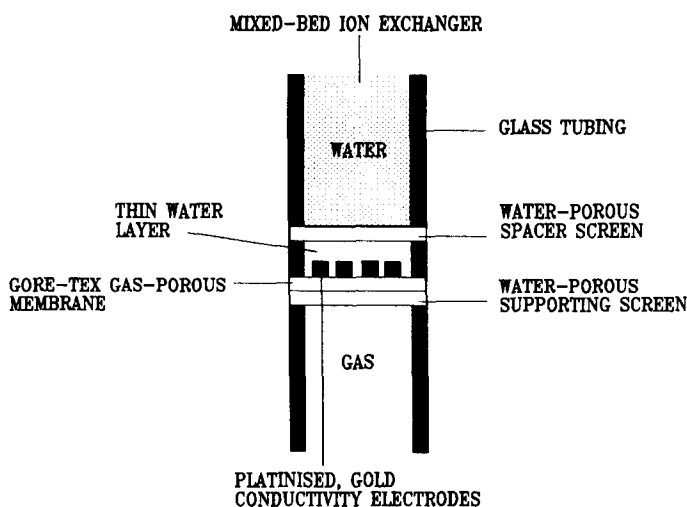


Fig. 6. Schematic representation of a continuous conductimetric CO<sub>2</sub> sensor. Electrodes positioned in thin water layer measure its conductance. Diffusion of CO<sub>2</sub> through the membrane into water layer and removal of ionic species through screen by ion exchanger establish a steady-state gradient of CO<sub>2</sub> [70, 71].

paramagnetically [27] or by means of the Clark-type amperometric electrode [43, 72] which consists typically of an Ag ring anode surrounding a Pt cathode (Fig. 7). When an appropriate potential is applied across the electrodes, O<sub>2</sub> is reduced at the cathode in the reaction:

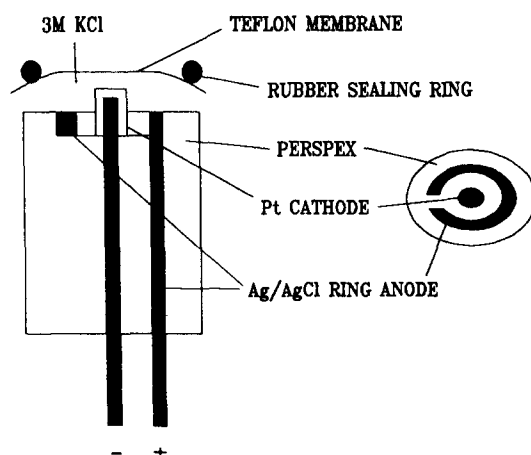
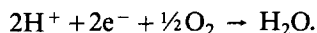
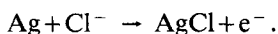


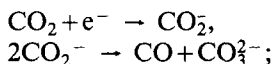
Fig. 7. Design of the Clark-type electrode. An Ag ring anode surrounds a Pt cathode and the circuit is completed by a KCl solution. Such electrodes are usually constructed in Perspex.

The circuit is completed by a KCl-salt bridge between the anode and the cathode and the anode reaction is:

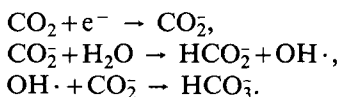


Thus, if  $\text{O}_2$  is present, a current flows and AgCl is deposited on the anode. As the potential of the cathode is made more negative,  $\text{O}_2$  is reduced more rapidly until a potential (difference) is reached where the reduction rate is equal to the rate at which  $\text{O}_2$  can diffuse to the cathode, producing a plateau region (diffusion-limited current) (Fig. 8) [73]. If the potential difference is increased beyond the plateau region the evolution of H commences at the cathode and there is a large increase in current. If less  $\text{O}_2$  is present its diffusion is slower and the plateau current is, therefore, lower. The amount of current is, thus, linear with the  $\text{O}_2$  concentration at the electrode surface.

$\text{CO}_2$  may also be reduced at such a cathode in the following way [74]:  
in anhydrous conditions,



in the presence of water,



Hence, an amperometric  $\text{CO}_2$  probe is, therefore, possible [75]. This electrode for amperometric  $\text{CO}_2$  measurement was a three-electrode system (Fig. 9), consisting of

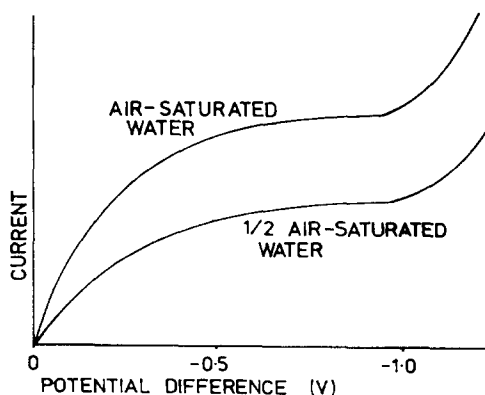


Fig. 8. Current-voltage curve for reduction of  $\text{O}_2$  in Clark-type amperometric electrode (Fig. 7). As potential difference between Pt cathode and Ag anode is increased, rate of  $\text{O}_2$  reduction at cathode increases. A plateau is reached when reduction rate of  $\text{O}_2$  is equal to rate at which  $\text{O}_2$  can diffuse to cathode. If potential difference is increased above plateau region, H evolution commences and there is a large increase in current.

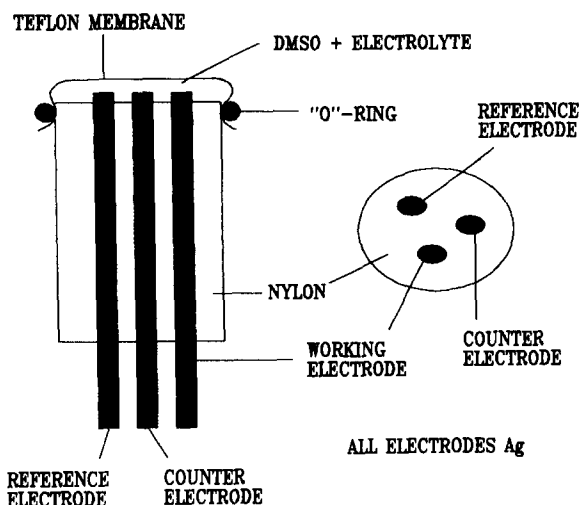


Fig. 9. Design of an amperometric  $\text{CO}_2$  electrode [102] based upon the design of an  $\text{O}_2/\text{CO}_2$  electrode by Albery and Barron [75]. Since Perspex dissolves in DMSO, the electrode was constructed from nylon.

a working electrode, a counter electrode and a reference electrode, as compared to the Clark-type electrode which is generally a two-electrode system (see Fig. 7). In such two-electrode systems, current passing through the reference electrode may cause its potential to deviate and give rise to internal polarisation. This, and resistive drops due to the presence of junction potentials across solutions and the interface between different materials, leads to a decrease in the absolute potential difference between the working electrode and the reference electrode. Addition of the third electrode minimises these problems as the current now flows between the working electrode and the counter electrode with the potential of the working electrode being set relative to that of the reference electrode [76–78].

To reduce the resistive drops to an insignificant level, inert supporting electrolytes are used [76, 79]. Potassium chloride is commonly used as a supporting electrolyte since it is easily available in high purity form and the mobility of the potassium ion and the chloride ion are almost exactly equal. Potassium nitrate is an alternative when chloride cannot be used. However, when a high concentration of potassium ions is present, the foot of its polarographic reduction wave may extend to values sufficiently positive as to cause unwanted interference [79]. This can be avoided by the use of tetraalkylammonium salts. Interference by the chloride ion often arises when mercury electrodes are used and may be avoided by using nitrate or perchlorate salts.

A number of processes has been reported for the reduction of  $\text{CO}_2$  although these have been directed primarily to aqueous solutions [e.g., 80–83]. However, at the potential required for the reduction of  $\text{CO}_2$  (at an Ag working electrode the  $\frac{1}{2}$ -wave potential is  $-2.13$  V vs. the standard calomel electrode [75]) water is electrolysed [84], thus, contra-indicating the use of aqueous solutions within the electrode for analytical purposes.  $\text{O}_2$  is much more readily reduced than is  $\text{CO}_2$  and, if present, will give rise to the production of superoxide which accumulates and interferes with the measurement

of  $p\text{CO}_2$  [75]. In the electrode described by Albery and Barron [75], this problem was circumvented by the use of a metallised membrane as an electrochemical filter for the removal of  $\text{O}_2$ . The principle of this is that  $\text{O}_2$  is reduced, at the metallised membrane in an aqueous solvent to produce water, at a lower potential than that which is required to reduce  $\text{CO}_2$ . However, in the case of an electrode intended for the monitoring of  $p\text{CO}_2$  in anaerobic fermentations the presence of  $\text{O}_2$  will not be a problem.

Determination of  $p\text{CO}_2$  with an amperometric electrode should be able to give continuous measurements, have a linear relationship between output and  $p\text{CO}_2$  (as compared with the Severinghaus-type electrode which has a logarithmic relationship between the variable actually measured, pH and  $p\text{CO}_2$  [50]) and be both simple and cheap to construct, especially, if it is to be used under anaerobic conditions.

Recently, a  $\text{CO}_2$  sensor using immobilised thermophilic bacteria and an amperometric  $\text{O}_2$  electrode was described [85]. The construction of this  $\text{CO}_2$  sensor is as shown in Fig. 10. The immobilised bacteria were supplied with  $\text{O}_2$ -saturated buffer containing various metal ions and  $200\ \mu\text{M}$  glucose.  $\text{CO}_2$  permeates through the dialysis membrane and is assimilated by the bacteria, causing their respiratory rate to increase [86]. The ' $\text{CO}_2$ ' concentration was estimated from the decrease in the output current from the  $\text{O}_2$  electrode, caused by the decrease in the  $\text{O}_2$  concentration corresponding to the increased bacterial respiration. Although the electrode produced an output with a linear relationship to ' $\text{CO}_2$ ', the range of  $\text{CO}_2$  concentrations tested was small, the temperature range was limited and the response time was 5–10 min.  $\text{CO}_2$  may be detected by another method which utilises an  $\text{O}_2$  electrode [87]. As with the immobilised bacterial sensor, the presence of  $\text{CO}_2$  results in a reduction, proportional to the  $p\text{CO}_2$ , of the current output of the  $\text{O}_2$  electrode.

**9.3.1. Reduction of  $\text{CO}_2$  mediated by an electrocatalyst.** Recently, a great deal of attention has been devoted to the utilisation of  $\text{CO}_2$  as a source of C in the elec-

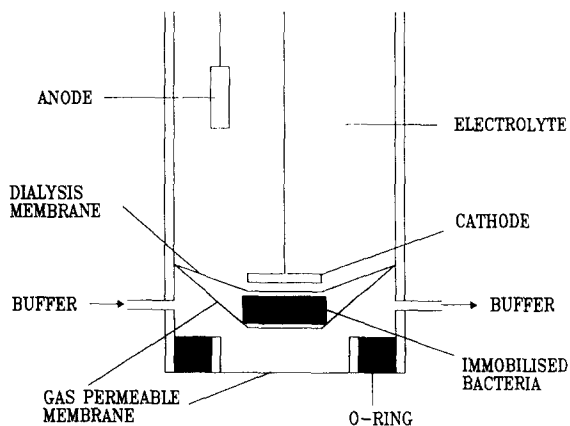
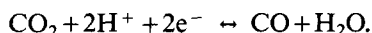


Fig. 10. Cross-section of a  $\text{CO}_2$  sensor which utilises immobilised thermophilic bacteria [85]. The bacteria assimilate  $\text{CO}_2$  and, in doing so, their respiration rate increases resulting in a decreased  $\text{O}_2$  concentration. It is a decrease in  $\text{O}_2$  concentration that is measured by the electrode. Such electrodes generally exhibit a rather poor dynamic range.

trochemical synthesis of organic products [e.g., 80, 88]. One possibility would be to use an electrocatalyst, developed for such CO<sub>2</sub> utilisation, in an amperometric CO<sub>2</sub> electrode, to reduce the potential required for the reduction of CO<sub>2</sub>. However, few catalytic systems are known despite the development of a number of different strategies (electro-, photo- and photo-electro-chemical). The purely photochemical systems may be classified as either heterogenous, making use of semiconductor suspensions [89–92], or homogenous, employing aqueous solutions of metal ions [82, 93–95], organic dyes or transition metal complexes [96–99]. In a system based on Re(bipy)(CO)<sub>3</sub>Cl, high yields of CO were photogenerated from CO<sub>2</sub> [100]. When Re(bipy)(CO)<sub>3</sub>Cl was used as a catalyst in the electrochemical reduction of CO<sub>2</sub> to CO [83], the system displayed high current efficiency and long-term stability (high overall turnover). Such catalytic properties would appear to make Re(bipy)(CO)<sub>3</sub>Cl an attractive catalyst for use in an amperometric CO<sub>2</sub> electrode for the improvement of selectivity and specificity. Direct noncatalysed reduction of CO<sub>2</sub> follows a monoelectronic pathway (equations given above), requiring potentials as negative as –2 V [74, 101]. On the other hand, polyelectronic reduction of CO<sub>2</sub> may occur at much less negative potentials, e.g., the E<sub>0</sub> for the dielectronic reduction of CO<sub>2</sub> to CO in aqueous solutions at pH 7 is only –0.52 V [83, 96] and occurs as follows:



Re(bipy)(CO)<sub>3</sub>Cl was reported to reduce CO<sub>2</sub> to CO at a potential of –1.25 V, substantially below the potential for monoelectronic CO<sub>2</sub> reduction.

Although the use of Re(bipy)(CO)<sub>3</sub>Cl in an electrode may in theory appear to be beneficial, in practice, this electrocatalyst offered no added benefit and, in fact, worsened matters by reducing the rate of current decay after a reduction in *p*CO<sub>2</sub> [102]. That the use of the electrocatalyst in an electrode caused an increase in the time taken for the current to decrease following a decrease in the *p*CO<sub>2</sub>, was possibly due to interaction between CO<sub>2</sub> and the electrocatalyst. Such interactions would ‘delay’ CO<sub>2</sub> desorption from the solvent giving rise to higher concentrations of ‘CO<sub>2</sub>’ than would normally be encountered and reflected in the increased current. However, the use of other electrocatalysts in amperometric electrodes may improve this method of CO<sub>2</sub> measurement.

## 10. Concluding Remarks and Summary

As with any set of analytical methods, there is likely to be a trade-off between resolution, precision, accuracy, response time, ease of use and expense. The methods discussed herein, and summarised in Table 2, cover the spectrum of simplicity. At present, mass spectrometry is probably the most widely used method industrially and/or where money is no object. Fibre optic probes seem to offer many advantages whilst continuing technological development work may be expected for potentiometric devices in particular. IR methods probably suffer too many interferences to be of general use in fermentations though the microwave region does not yet seem to have been explored.

TABLE 2

SUMMARY OF METHODS FOR ESTIMATION OF 'CO<sub>2</sub>' DESCRIBED IN TEXT

Method	Applications	Comments
Titrimetric	Liquid samples	Unsuitable for continuous use
Colourimetric	Liquid or gaseous samples	Unsuitable for continuous use
Gravimetric	Exhaust gases	Unsuitable for continuous use
Volumetric	Dissolved 'CO <sub>2</sub> '	Not able to provide genuinely continuous monitoring. Cumbersome
Enzymatic	Dissolved 'CO <sub>2</sub> '	Specific for 'CO <sub>2</sub> ' but unsuitable for continuous use
Katharometer	Detection of gases	Not specific for CO <sub>2</sub> . Should ideally be combined with a separation method, e.g., GC
Mass spectrometry	Dissolved gases or fermentor exhaust gases	Expensive but precise and accurate
IR	Exhaust gases	Expensive. Not specific for CO <sub>2</sub>
Fibre optic probes	Dissolved 'CO <sub>2</sub> ' Overcomes problems of making miniature glass electrodes	Measures pH – organic acids or bases may interfere
Gas chromatography	Exhaust gas or small liquid samples	Unsuitable for continuous use
Potentiometric	Dissolved 'CO <sub>2</sub> '	Slow response time, electrodes not cheap, organic acids/bases may interfere, logarithmic response
Conductimetric	Exhaust gases	Evaluated only up to 0.01 atm
Amperometric	Exhaust gases	Simple and cheap.
Piezoelectric	Exhaust gases	Simple and cheap. Interferences not known but volatile interferences likely

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**Noted added in proof**

Fatibello-Filho et al. [103] have recently described a CO<sub>2</sub>-sensitive piezoelectric crystal which gave a linear response to *p*CO<sub>2</sub> in the range of 0.018–0.16 atm under fermentation conditions.

**References**

- 1 Knoche, W. (1980) Chemical reactions of CO<sub>2</sub> in water. In: *Biophysics and Physiology of Carbon Dioxide* (Bauer, C., Gros, G. and Bartels, H., eds.), pp. 3–11, Springer-Verlag, Berlin.
- 2 Covington, A. K. (1985) Potentiometric titrations of aqueous carbonate solutions. *Chem. Soc. Rev.* 14, 265–281.
- 3 Yagi, H. and Yoshida, F. (1977) Desorption of CO<sub>2</sub> from fermentation broth. *Biotechnol. Bioeng.* 19, 801–819.

- 4 Butler, J.N. (1982) CO<sub>2</sub> Equilibria and Their Applications, Addison Wesley, London.
- 5 Schumpe, A., Quicker, G. and Deckwer, W.-D. (1982) Gas solubilities in microbial culture media. *Adv. Biochem. Eng.* 24, 1–38.
- 6 Ho, C.S., Smith, M.D. and Shanahan, J.F. (1987) Carbon dioxide transfer in biochemical reactors. *Adv. Biochem. Eng. Biotechnol.* 35, 83–125.
- 7 Pirt, S.J. (1975) *Principles of Microbe and Cell Cultivation*, Blackwell Scientific Publications, Oxford.
- 7a Canongate Technology Ltd (1983) The diffusion of gases through a silicone rubber membrane, and its application to an in-line carbonation meter. *Proc. Meet. MBAA San Diego*.
- 8 Hayward, A.C. (1957) Detection of gas production from glucose by heterofermentative lactic acid bacteria. *J. Gen. Microbiol.* 16, 9–15.
- 9 Gibson, T. and Abdel-Malek, Y. (1945) The formation of carbon dioxide by lactic acid bacteria and *Bacillus licheniformis* and a cultural method of detecting the process. *J. Dairy Res.* 14, 35–44.
- 10 Rogosa, M., Wiseman, R.F., Mitchell, J.A., Disraely, M.N. and Beauman, A.J. (1953) Species differentiation of oral lactobacilli from man including descriptions of *Lactobacillus cellobiosus* nov. spec. *J. Bacteriol.* 65, 681–699.
- 11 Eldredge, E.E. and Rogers, L.A. (1914) The bacteriology of cheese of the Emmenthal type. *Zentralbl. Bakteriologie. Parasitenkd. Infektionskr. Hyg. Abt. 2.* 40, 5–21.
- 12 Williams, O.B. and Campbell, L.L. (1952) The detection of heterofermentation by lactic acid bacteria. *Food Technol.* 5, 306.
- 13 Hammer, B.W. and Baker, M.P. (1923) Studies on *Streptococcus paracitrovorus* group. *Iowa Agric. Home Econ. Exp. Stn. Res. Bull.* 81.
- 14 Sperber, W.H. and Swan, J. (1976) Hot-loop test for the determination of carbon dioxide production from glucose by lactic acid bacteria. *Appl. Environ. Microbiol.* 31, 990–991.
- 15 Dicks, L.M.T. and van Vuuren, H.J.J. (1987) A modification of the hot-tube method for the detection of carbon dioxide produced by heterofermentative *Lactobacillus* strains. *J. Microbiol. Methods.* 6, 273–275.
- 16 Milton, R.F. (1955) Illustrative examples of microvolumetric procedures. In: *Methods of Quantitative Micro-Analysis* (Milton, R.F. and Waters, W.A., eds.), pp. 170–224, Edward Arnold, London.
- 17 Conway, E.J. (1962) *Microdiffusion Analysis and Volumetric Error*, Crosby Lockwood & Son, London.
- 18 Obrink, K.J. (1955) A modified Conway unit for microdiffusion analysis. *Biochem. J.* 59, 134–136.
- 19 Greenberg, A.E., Connors, J.J., Jenkins, D. and Franson, M.A.H. (1981) *Standard Methods for the Examination of Water and Wastewater*, APHA-AWWA-WPCF, Washington.
- 20 Vogel, A.I. (1959) *A Text-Book of Quantitative Inorganic Analysis Theory and Practice*, Longmans, Green & Co., London.
- 21 Milton, R.F. (1955) Colorimetric analysis. In: *Methods of Quantitative Micro-Analysis* (Milton, R.F. and Waters, W.A., eds.), pp. 229–420, Edward Arnold, London.
- 22 Snell, F.D. and Snell, C.T. (1959) *Colorimetric Methods of Analysis*, Vol. IIA, D. Van Nostrand Co., Princeton, New Jersey.
- 23 Rozzi, A., Burton, K.W. and Hawkes, D.L. (1983) Potentiometric method for the determination of carbon dioxide in biogas. *J. Agric. Eng. Res.* 28, 505–512.
- 24 Snell, F.D. and Snell, C.T. (1961) *Colorimetric Methods of Analysis*, Vol. II, D. Van Nostrand Co., Princeton, New Jersey.
- 25 Maxon, W.D. and Johnson, M.J. (1952) Continuous photometric determination of carbon dioxide in gas streams. *Anal. Chem.* 24, 541–545.
- 26 Wilson, K.M. (1955) Gasometric methods of micro-analysis. In: *Methods of Quantitative Micro-Analysis* (Milton, R.F. and Water, W.A., eds.), pp. 521–605, Edward Arnold, London.
- 27 Elsworth, R. (1970) The measurement of oxygen absorption and carbon dioxide evolution in stirred deep cultures. In: *Methods in Microbiology 2* (Norris, J.R. and Ribbons, D.W., eds.), pp. 213–228, Academic Press, London.
- 28 Van Slyke, D.D. and Neill, J.M. (1924) The determination of gases in blood and other solutions by vacuum extraction and manometric measurement. *J. Biol. Chem.* 61, 523–573.
- 29 Dixon, M. (1952) *Manometric Methods as Applied to the Measurement of Cell Respiration and Other Processes*, Third Edition, Cambridge University Press, Cambridge.
- 30 Umbreit, W.W., Burris, R.H. and Stauffer, J.F. (1957) *Manometric Techniques*, Burgess Publishing, Minneapolis, Minnesota.

- 31 Rozzi, A. and Labellarte, G. (1984) Direct bicarbonate determination in anaerobic digester liquors by measurement of the pressure of carbon dioxide. *Proc. Biochem.* 19, 201–203.
- 32 van Stekelenburg, G. V., Valk, C. and van Wijngaarden-Penterman (1985) An extremely simple and fast microlitre method for the determination of total carbon dioxide in biological samples. *Ann. Clin. Biochem.* 22, 509–513.
- 33 Forrester, R. L., Wataji, L. J., Silverman, D. A. and Pierre, K. J. (1976) Enzymatic method for determination of  $\text{CO}_2$  in serum. *Clin. Chem.* 22, 243–245.
- 34 Panteghini, M., Calarco, M., Malchiodi, A. and Bonora, R. (1985) Direct potentiometric analysis of sodium, potassium, chloride, and total carbon dioxide in serum. *Int. Clin. Prod. Rev.* 4, 28–35.
- 35 Odham, G. and Larsson, L. (1984) Mass spectrometry. In: *Gas Chromatography/Mass Spectrometry Applications in Microbiology* (Odham, G., Larsson, L. and Mårdh, P.-A., eds.), pp. 27–54, Plenum Press, New York.
- 36 Larsson, L. and Odham, G. (1984) Gas chromatography. In: *Gas Chromatography/Mass Spectrometry Applications in Microbiology* (Odham, G., Larsson, L. and Mårdh, P.-A., eds.), pp. 7–26, Plenum Press, New York.
- 37 Fleischaker, R. J., Weaver, J. C. and Sinskey, A. (1981) Instrumentation for process control in cell culture. *Adv. Appl. Microbiol.* 27, 137–167.
- 38 Lloyd, D., Scott, R. I. and Williams, T. N. (1983) Membrane inlet mass spectrometry – measurement of dissolved gases in fermentation liquids. *Trends Biotechnol.* 1, 60–63.
- 39 Lloyd, D., Bohátka, S. and Szilágyi, J. (1985) Quadrupole mass spectrometry in the monitoring and control of fermentations. *Biosensors* 1, 179–212.
- 40 Lloyd, D. and Scott, R. I. (1983) Direct measurement of dissolved gases in microbiological systems using membrane inlet mass spectrometry. *J. Microbiol. Methods* 1, 313–328.
- 41 Bohátka, S., Langer, G., Szilágyi, J. and Berecz, I. (1983) Gas concentration determination in fermentors with quadrupole mass spectrometer. *Int. J. Mass Spectrom. Ion Phys.* 48, 277–280.
- 42 Heinzle, E., Bolzern, O., Dunn, I. J. and Bourne, J. R. (1981) A porous membrane-carrier gas measurement system for dissolved gases and volatiles in fermentation systems. In: *Advances in Biotechnology, Vol. 1, Scientific and Engineering Principles* (Moo-Young, M., Robinson, C. W. and Vezina, C., eds.), pp. 439–444, Pergamon Press, Oxford.
- 43 Tebbutt, P., Clark, D., Robinson, G., Hahn, C. E. W. and Alberly, W. J. (1986) The electrochemistry of gases of medical interest. In: *Electrochemistry, Sensors and Analysis* (Smyth, M. R. and Vos, J. G., eds.), pp. 315–322, Elsevier, Amsterdam.
- 44 Telling, R. C., Elsworth, R. and East, D. N. (1958) A continuous infrared analyser for measurement of  $\text{CO}_2$  in effluent air from bacterial cultures. *J. Appl. Bacteriol.* 21, 26–44.
- 45 Maxwell, J. C. and Caughey, W. S. (1978) Infrared spectroscopy of ligands, gases, and other groups in aqueous solutions and tissue. *Methods Enzymol.* 54, 302–323.
- 46 Peterson, J. I., Goldstein, S. R., Fitzgerald, R. V. and Buckhold, D. K. (1980) Fiber optic pH probe for physiological use. *Anal. Chem.* 52, 864–869.
- 47 Vurek, G. G., Peterson, J. I., Goldstein, S. R. and Severinghaus, J. W. (1982) Fiber optic  $p\text{CO}_2$  probe. *Fed. Proc. Fed. Am. Soc. Exp. Biol.* 41, 1483.
- 48 Vurek, G. G., Feustel, P. J. and Severinghaus, J. W. (1983) A fiber optic  $p\text{CO}_2$  sensor. *Ann. Biomed. Eng.* 11, 499–510.
- 49 Coon, R. L., Lai, N. C. J. and Kampine, J. P. (1976) Evaluation of a dual-function pH and  $p\text{CO}_2$  in vivo sensor. *J. Appl. Physiol.* 40, 625–629.
- 50 Nicholls, D. G. and Garland, P. B. (1972) Electrode measurements of carbon dioxide. In: *Methods in Microbiology* 6b (Norris, J. R. and Ribbons, D. W., eds.), pp. 55–63, Academic Press, London.
- 51 Fogt, E. J., Untereker, D. F., Norenberg, M. S. and Meyerhoff, M. E. (1985) Response of ion-selective field effect transistors to carbon dioxide and organic acids. *Anal. Chem.* 57, 1995–1998.
- 52 Perry, J. A. (1981) *Introduction to Analytical Gas Chromatography*, Marcel Dekker, New York.
- 53 Grob, R. L. (1977) *Modern Practice of Gas Chromatography*, John Wiley & Sons, New York.
- 54 Cramers, C. A. and McNair, H. M. (1983) Gas chromatography. *Chromatogr. J. Chromatogr. Libr.* 22a, A195–A224.
- 55 Willett, J. E. (1987) *Gas Chromatography*, John Wiley & Sons, New York.
- 56 Kell, D. B. (1980) The role of ion-selective electrodes in improving fermentation yields. *Proc. Biochem. Jan.*, 1–6.
- 57 Clarke, D. J., Kell, D. B., Morris, J. G. and Burns, A. (1982) The role of ion-selective electrodes in

- microbial process control. *Ion-Sele. Electr. Rev.* 4, 75–131.
- 58 Clarke, D.J., Calder, M.R., Carr, R.J.G., Blake-Coleman, B.C., Moody, S.C. and Collinge, T.A. (1985) The development and application of biosensing devices for bioreactor monitoring and control. *Biosensors* 1, 213–320.
  - 59 Donaldson, T.L. and Palmer, H.J. (1979) Dynamic response of the carbon dioxide electrode. *Am. Inst. Chem. Eng.* 25, 143–151.
  - 60 van der Schoot, B. and Bergveld, P. (1984) Prediction of the dynamic response of the potentiometric carbon dioxide electrode. *Anal. Chim. Acta* 166, 93–101.
  - 61 Puhar, E., Einsele, A., Buhler, H. and Ingold, W. (1980) Steam-sterilisable  $p\text{CO}_2$  electrode. *Biotechnol. Bioeng.* 22, 2411–2416.
  - 62 Gauthier, M., Belanger, A. and Fauteux, D. (1983) Solid carbonate electrolytes for the potentiometric measurement of carbon oxides and carbon bearing materials. In: *Chemical Sensors* (Seiyama, T., Fueki, K., Shiokawa, J. and Suzuki, S., eds.), pp. 353–356, Elsevier, Amsterdam.
  - 63 Cheung, P.W., Neuman, M.R., Fleming, D.G. and Ko, W.H. (1978) *Theory, Design, and Biomedical Applications of Solid State Chemical Sensors*, CRC Press, West Palm Beach.
  - 64 Covington, A.K. and Sibbald, A. (1987) Ion-selective field-effect transistors (ISFETs). *Phil. Trans. R. Soc. London Ser. B*, 316, 31–46.
  - 65 van den Berg, A. (1988) Ion Sensors Based on ISFET's with Synthetic Ionophores. Ph.D. thesis. Centrum voor Micro-Elektronica, Enschede.
  - 66 Alford, J.S. (1976) Measurement of dissolved  $\text{CO}_2$ . *Can J. Microbiol.* 22, 52–56.
  - 67 Hazinski, T.A. and Severinghaus, J.W. (1982) Transcutaneous analysis of arterial  $p\text{CO}_2$ . *Med. Instrumen.* (Baltimore) 16, 150–153.
  - 68 Hagihara, B., Fujiwara, Y., Ohkawa, S., Yotsuya, K., Hasegawa, T., Shimizu, K. and Kurachi, K. (1983) Transcutaneous  $\text{CO}_2$  electrode. In: *Chemical Sensors* (Seiyama, T., Fueki, K., Shiokawa, J. and Suzuki, S., eds.), pp. 585–590, Elsevier, Amsterdam.
  - 69 Mendelson, Y. and Peura, R.A. (1984) Noninvasive transcutaneous monitoring of arterial blood gases. *IE Trans. Biomed. Eng.* 31, 792–800.
  - 70 Bruckenstein, S. and Symanski, J.S. (1986) Continuous conductometric sensor for carbon dioxide. *Anal. Chem.* 58, 1766–1770.
  - 71 Bruckenstein, S. and Symanski, J.S. (1986) Analytical applications of gas membrane electrodes. *J. Chem. Soc. Faraday Trans. 1* 82, 1105–1116.
  - 72 Fatt, I. (1976) *Polarographic Oxygen Sensors*, CRC Press, New York.
  - 73 Beechey, R.W. and Ribbons, D.W. (1972) Oxygen electrode measurements. In: *Methods in Microbiology*, Vol. 6b (Norris, J.R. and Ribbons, D.W., eds.), pp. 25–53, Academic Press, London.
  - 74 Haynes, L.V. and Sawyer, D.T. (1967) Electrochemistry of carbon dioxide in dimethyl sulfoxide at gold and mercury electrodes. *Anal. Chem.* 39, 332–338.
  - 75 Albery, W.J. and Barron, P. (1982) A membrane electrode for the determination of  $\text{CO}_2$  and  $\text{O}_2$ . *J. Electroanal. Chem.* 138, 79–87.
  - 76 Sawyer, D.T. and Roberts, J.L. (1974) *Experimental Electrochemistry for Chemists*. Wiley-Interscience, London.
  - 77 Bond, A.M. (1980) *Modern Polarographic Methods in Analytical Chemistry*, Marcel Dekker, New York.
  - 78 Kuhn, A.T. (1987) Electrochemical techniques. In: *Techniques in Electrochemistry, Corrosion and Metal Finishing – A Handbook* (Kuhn A.T., ed.), pp. 55–74, John Wiley & Sons, New York.
  - 79 Plambeck, J.A. (1982) *Electroanalytical Chemistry. Basic Principles and Applications*, John Wiley & Sons, New York.
  - 80 Bennett, E.M., Eggins, B.R., McNeill, J. and McMullan, E.A. (1980) Recycling carbon dioxide from fossil fuel combustion. *Anal. Proc.* 17, 356–359.
  - 81 Eggins, B.R. and McNeill, J. (1983) Voltammetry of carbon dioxide. Part I. A general survey of voltammetry at different electrode materials in different solvents. *J. Electroanal. Chem.* 148, 17–24.
  - 82 Tinnemans, A.H.A., Koster, T.P.M., Thewissen, D.H.M.W. and Mackor, A. (1983) Photoassisted reduction of  $\text{CO}_2$  over aqueous suspensions of strontium titanate powders and electrolytic reduction of  $\text{CO}_2$  at titanate electrodes. In: *Photochemical, Photoelectrochemical and Photobiological Processes* (Hall, D.O., Palz, W. and Pirrwitz, D., eds.), Sol. Energy R D Eur. Community Ser. D 2, 86–91.
  - 83 Hawecker, J., Lehn, J.-M. and Ziessel, R. (1984) Electrocatalytic reduction of carbon dioxide mediated by  $\text{Re}(\text{bipy})(\text{CO})_3\text{Cl}$  (bipy = 2,2'-bipyridine). *J. Chem. Soc. Chem. Comm.* 328–330.

- 84 Chang, R. (1981) *Physical Chemistry with Applications to Biological Systems*, Collier Macmillan Publishers, London.
- 85 Suzuki, H., Tamiya, E., Karube, I. and Oshima, T. (1988) Carbon dioxide sensor using thermophilic bacteria. *Anal. Lett.* 21, 1323–1336.
- 86 Karube, I., Tamiya, E., Sode, K., Yokoyama, K., Kitagawa, Y., Suzuki, H. and Asano, Y. (1988) Application of microbiological sensors in fermentation processes. *Anal. Chim. Acta* 213, 69–77.
- 87 Mills, A. and Lawrence, C. (1985) Determination of electroactive and non-electroactive gases using a membrane polarographic detector in a flow system. *Analyst* 110, 23–26.
- 88 Aresta, M. and Forti, G. (1987) Carbon Dioxide as a Source of Carbon. *Biochemical and Chemical Uses*. D. Reidel Publishing Co., Dordrecht.
- 89 Inoue, T., Fujishima, A., Konishi, S. and Honda, K. (1979) Photoelectrocatalytic reduction of carbon dioxide in aqueous suspensions of semiconductor powders. *Nature (London)* 277, 637–638.
- 90 Taniguchi, I., Aurian-Blajeni, B. and Bockris, O'M. (1983) Photo-aided reduction of carbon dioxide to carbon monoxide. *J. Electroanal. Chem.* 157, 179–182.
- 91 Ito, K., Ikeda, S., Ohta, S. and Iida, T. (1984) On the reduction products of carbon dioxide at a *p*-type gallium phosphide photocathode in aqueous electrolytes. *Bull. Chem. Soc. Jpn.* 57, 583–584.
- 92 Ikeda, S., Yoshida, M. and Ito, K. (1985) Photoelectrochemical reduction products of carbon dioxide at metal coated *p*-GaP photocathodes in aqueous electrolytes. *Bull. Chem. Soc. Jpn.* 58, 1353–1357.
- 93 Tinnemans, A. H. A., Koster, T. P. M., Thewissen, D. H. M. W. and Mackor, A. (1982) Formation of methanol and other C1-C3 compounds in the photoassisted reaction of formaldehyde and water over strontium titanate suspensions containing transition metal oxide deposits. *Nouv. J. Chim.* 6, 373–379.
- 94 Ulman, M., Tinnemans, A. H. A., Mackor, A., Aurian-Blajeni, B. and Halmann, M. (1982) Photoreduction of carbon dioxide to formic acid, formaldehyde, methanol, acetaldehyde and ethanol using suspensions of strontium titanate with transition metal additives. *Int. J. Sol. Energy* 1, 213–222.
- 95 Thampi, K. R., Kiwi, J. and Gratzel, M. (1987) Methanation and photo-methanation of carbon dioxide at room temperature and atmospheric pressure. *Nature (London)* 327, 506–508.
- 96 Fisher, B. and Eisenberg, R. (1980) Electrocatalytic reduction of carbon dioxide by using macrocycles of nickel and cobalt. *J. Am. Chem. Soc.* 102, 7361–7363.
- 97 Lehn, J.-M. and Ziessel, R. (1982) Photochemical generation of carbon monoxide and hydrogen by reduction of carbon dioxide and water under visible light irradiation. *Proc. Natl. Acad. Sci. U.S.A.* 79, 701–704.
- 98 Tinnemans, A. H. A., Koster, T. P. M., Thewissen, D. H. M. W. and Mackor, A. (1984) Tetraaza-macrocylic cobalt (II) and nickel (II) complexes as electron-transfer agents in the photo(electro)chemical and electrochemical reduction of carbon dioxide. *J. R. Neth. Chem. Soc.* 103, 288–295.
- 99 Gambarotta, S., Strologo, S., Floriani, C., Chiesi-Villa, A. and Guastini, C. (1985) Stepwise reduction of carbon dioxide to formaldehyde and methanol: reactions of CO<sub>2</sub> and CO<sub>2</sub>-like molecules with hydrido-chlorobis(cyclopentadienyl)zirconium (IV). *J. Am. Chem. Soc.* 107, 6278–6282.
- 100 Hawecker, J., Lehn, J.-M. and Ziessel, R. (1983) Efficient photochemical reduction of CO<sub>2</sub> to CO by visible light irradiation of systems containing Re(bipy)(CO)<sub>3</sub>X or Ru(bipy)<sub>3</sub><sup>2+</sup> – Co<sup>2+</sup> combinations as homogenous catalysts. *J. Chem. Soc., Chem. Comm.* 536–538.
- 101 Roberts, J. L. and Sawyer, D. T. (1965) Voltammetric determination of carbon dioxide using dimethyl-sulfoxide as a solvent. *J. Electroanal. Chem.* 9, 1–7.
- 102 Dixon, N. M. (1988) Effects of CO<sub>2</sub> on anaerobic bacterial growth and metabolism. Ph.D. Thesis, University College of Wales, Aberystwyth.
- 103 Fatillo-Filho, O., de Andrade, J. F., Suleiman, A. A. and Guilbault, G. G. (1989) Piezoelectric crystal monitor for carbon dioxide in fermentation processes. *Anal. Chem.* 61, 746–748.